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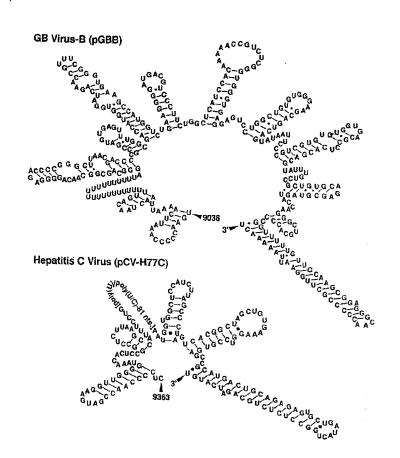
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(54) Title: INFECTIOUS cDNA CLONE OF GB VIRUS B AND USES THEREOF



(57) Abstract: The present invention relates to nucleic acid sequence which comprises the genome of an infectious GB virus B clone. The invention also relates to the use of the nucleic acid sequence of the infectious GB virus B clone to indirectly study the molecular properties of HCV, and in the production of HCV/GBV-B chimeras. The invention further relates to the use of the infectious nucleic acid sequence of GB virus B clone and the HCV/GBV-B chimeras in the development of vaccines and therapeutics for HCV.

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Title of Invention

Infectious cDNA clone of GB Virus B and Uses Thereof

Field of Invention

sequence which comprises the genome of an infectious GB virus B (GBV-B) clone. The invention also relates to the use of the nucleic acid sequence of the infectious GB virus B clone to study indirectly the molecular properties of hepatitis C virus (HCV), and in the production of HCV/GBV-B chimeras. The invention further relates to the use of the infectious nucleic acid sequence of the GB virus B clone and the HCV/GBV-B chimeras in the development of vaccines and therapeutics for HCV.

Background of Invention

Transmission studies of potential human hepatitis agents were first reported in 1967 (Deinhardt 1967). Four tamarins inoculated with acute phase sera from a surgeon with acute hepatitis (patient GB) developed hepatitis, as did most tamarins inoculated in serial passage studies. Subsequent studies indicated that the etiological agent responsible for the development of hepatitis in these animals was not any of the known human hepatitis viruses (Purcell 1994). In 1995, two related RNA viruses named GB virus-B (GBV-B) and GB virus A (GBV-A) were identified in acute phase sera of a tamarin which developed hepatitis following inoculation with serum of the eleventh tamarin passage of the putative GB agent (Simons 1995a).

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GBV-B infection of tamarins resulted in acute resolving hepatitis (Schlauder 1995, Buhk 1997). The

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natural host of GBV-B is still unknown as the virus has not been detected in uninoculated animals or in humans.

GBV-A, on the other hand, is an indigenous tamarin virus rather than a component of the original GB inoculum (Bukh 1997, Erker 1998). Experimental infection of tamarins with GBV-A did not produce hepatitis (Schlauder 1995). A human agent, GBV-C or hepatitis G virus, most closely related to GBV-A, was later identified (Simons 1995b, Linnen 1996). However, it is still not clear whether this virus actually causes hepatitis (Alter 1998, Bukh 1998a). Thus, of the known GB viruses, GBV-B may be the only true hepatitis virus.

Based on analysis of their genomic sequences, GBV-A, GBV-B and GBV-C were classified as members of the Flaviviridae family of viruses, and among the known viruses, GBV-B is the virus most closely related to hepatitis C virus (HCV) (Muerhoff 1995, Robertson 1998).

The GBV-B virus contains a positive-sense, single-stranded RNA genome of 9143 nucleotides (nts) (Simons 1995a, Muerhoff 1995). The viral genome of GBV-B consists of a 5' untranslated region (UTR), a single long open reading frame (ORF) and a 3' UTR. Based on known motifs, structural proteins were predicted to be encoded in the 5' portion of the ORF and nonstructural (NS) proteins in the 3' portion of the ORF (Muerhoff 1995). The hydropathy plots of the polyproteins of GBV-B and HCV are very similar even though the overall homology of the predicted polyproteins between GBV-B and HCV is only about 25-30% (Muerhoff 1995). The putative envelope proteins (E1 and E2) of GBV-B and HCV share common structural features, and significant homology was observed between the NS3 serine protease, the NS3 RNA

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helicase, and the NS5 RNA-dependent RNA polymerase regions of GBV-B and HCV (Muerhoff 1995). Furthermore, the function and substrate specificity of the GBV-B and HCV NS3 serine proteases are also similar (Scarselli 1997). The genomic structure and organization of GBV-B 5 and HCV share additional features of interest. First, colinear regions with significant sequence homology were identified in the 5' UTRs (Muerhoff 1995) and the predicted IRES structure of GBV-B is similar to that of 10 HCV (Lemon 1997). Second, both viruses begin the 3' UTR with a short sequence followed by a poly (U) stretch followed by additional nucleotides (50 nucleotides for GBV-B and 98 nucleotides for HCV). However, the 3' terminal sequence of HCV forms a stable stem-loop 15 structure (Kolykhalov 1996) whereas the published 3' terminal sequence of GBV-B does not.

To date, molecular studies of HCV are severely limited by the lack of an efficient cell culture system for the virus and by expense and limited availability of chimpanzees, the sole animal model for HCV.

Accordingly, a less expensive and more readily available animal than chimpanzees is necessary as an animal model for the study of HCV.

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Summary of Invention

The present invention relates to nucleic acid sequence which comprises the genome of an infectious GB virus B (GBV-B) clone. It is therefore an object of the invention to provide nucleic acid sequence which encodes an infectious GBV-B. Such nucleic acid sequence is referred to throughout the application as "infectious nucleic acid sequence".

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As significant structural homology exists between the genomes of GBV-B and HCV, the invention also relates to the use of infection of tamarins with the infectious nucleic acid sequence of GBV-B or with mutants of the infectious sequence to study indirectly the molecular properties of hepatitis C virus (HCV) or as a preliminary screen to identify agents which have antiviral activity against HCV.

nucleic acid sequences" consisting of portions of the infectious nucleic acid sequence of GBV-B and portions of the nucleic acid sequences of other viruses closely related to GBV-B such as HCV, GBV-C or other members of the Flaviviridae family which do not replicate in tamarins. In a preferred embodiment, the chimeric nucleic acid sequences consist of portions of the infectious nucleic acid sequence of GBV-B and portions of the nucleic acid sequence of HCV. The nucleic acid sequences taken from GBV-B and HCV can be open-reading frame sequences, and/or sequences from the 5'UTR and/or 3'UTR.

In one embodiment, GBV-B/HCV chimeras may be made in which 5' or 3' UTR sequences of the GBV-B infectious clone are replaced with the corresponding sequence from an HCV clone.

In another embodiment, GBV-B/HCV chimeras may be constructed in which the structural or non-structural regions of GBV-B are replaced by corresponding regions of HCV. Thus, such a chimera would contain, for example, the HCV structual region in a GBV-B "genomic backbone". Of course, it is understood by one of skill in the art that the construction of the above-described

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chimeric nucleic acid sequences may be reversed such that, for example, the GBV structural region may replace the structual region of an HCV genome to produce a chimera in which the GBV structural region is contained in an HCV backbone.

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The invention further relates to the use of the chimeric nucleic acid sequences of the invention to study the functions of HCV genes, and for the development of vaccine and antiviral agents against HCV.

The invention also relates to the use of the infectious GBV-B nucleic acid sequence, the mutated GBV-B nucleic acid sequences or the chimeric sequences of the invention to identify cell lines capable of supporting the replication of GBV-B or the chimeras of the invention.

The present invention also relates to the polypeptides encoded by the nucleic acid sequences of the invention or fragments thereof.

The present invention further relates to the <u>in vitro</u> and <u>in vivo</u> production of GBV-B, mutant GBV-B viruses or chimeric GBV-B/HCV viruses from the nucleic acid sequences of the invention.

The invention also provides that the nucleic acid sequences and viruses of the invention be supplied in the form of a kit, alone or in the form of a pharmaceutical composition.

Brief Description Of Figures

Figure 1 shows a flow diagram of GB virus transmission studies in two species of tamarins, Saguinus mystax (SM) and Saguinus oedipus (SO). The animals infected with GBV-B (Simons 1995a) are boxed.

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Two serum pools (GB 8/93 and GB 2/94) were made from acutely infected animals. Both pools contained GBV-B, as well as GBV-A (Simons 1995) at a titer of 10⁸ genome equivalent (GE)/ml. A 10% liver homogenate (CT 11/91) was made from a sacrificed tamarin. A number of S. mystax tamarins (SM 737, 749, 750, 760, 782, 795 and 799) and S. oedipus tamarins (SO 100) were naturally infected with GBV-A_{SM} and GBV-A_{SO}, respectively, prior to inoculation (Bukh 1997). Only two tamarins (SM 720 and 748), both GBV-A_{SM} negative, became infected with GBV-A (Simons 1995) following inoculation. Tamarins SM42 and SM670 were not tested for GBV-A or GBV-A_{SM}.

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Figure 2 shows the course of GBV-B infection in tamarins (S. mystax) inoculated with a dilution series of the GB 2/94 pool. All animals were inoculated intravenously at week 0 with 1 ml of the indicated dilution. Results of qualitative RT-nested PCR for GBV-B in serum are shown at the top (filled circles, positive; empty circles, negative). Serum levels of isocitrate dehydrogenase (ICD in units/ml); shaded area) and the estimated log10 GBV-B GE titer (vertical columns) were plotted against time.

Figure 3 shows alignment of the 3' UTR sequences of GBV-B. The sequence of the infectious clone of GBV-B (pGBB) is shown at the top (nts. 9038-9399).

The other sequences shown are: pGBB5-1, a non-infectious clone of GBV-B; GBV-B, a prototype of GBV-B (Simons 1995); eleven "gb" clones obtained from CT 11/91 liver homogenate by 5' RACE on the minus-strand GBV-B RNA; four "29" clones obtained from GB 2/94 pool by RT-PCR across 5'-to-3'-end-ligated viral GBV-B RNA; and seven "GBB3" clones obtained from GB 2/94 pool by standard RT-PCR.

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With pGBB as the reference, nucleotide substitutions or insertions are shown as uppercase letters, identical nucleotides are shown as dots and nucleotide deletions are shown as dashes.

Figure 4 shows the predicted secondary structure of the 3' UTRs of GBV-B and HCV as determined by the program "mfold" (Genetics Computer Group).

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Figure 5 shows the course of GBV-B infection in S. mystax tamarins transfected with RNA transcripts of pGBB. Both animals were negative for GBV-A_{SM}. At week 0 transcription mixtures were injected into tamarins by percutaneous intrahepatic injection guided by ultrasound. Results of qualitative RT-nested PCR for GBV-B in serum is shown at the top (filled circles, positive; empty circles, negative). Serum levels of isocitrate dehydrogenase (ICD in units/ml; shaded area) and the estimated log₁₀ GBV-B GE titer (vertical columns) were plotted against time.

Figures 6A-6F show the nucleotide sequence of the infectious hepatitis C virus clone of genotype 1a strain H77C and Figures 6G-6H show the amino acid sequence encoded by the clone.

Figures 7A-7F show the nucleotide sequence of the infectious hepatitis C virus clone of genotype 1b strain HC-J4 and Figures 7G-H show the amino acid sequence encoded by the clone.

Description of The Invention

The present invention relates to nucleic acid sequence which comprises the genome of an infectious GB virus B (GBV-B) clone. The nucleic acid sequence which comprises the genome of an infectious GBV-B virus is

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shown in SEQ ID NO:1 and is contained in the plasmid construct pGBB deposited with the American Type Culture Collection (ATCC) on May 28, 1999 and having ATCC accession number PTA-152. The present invention relates to the identification of a 260 nucleotide sequence at the 3' end of the infectious GBV-B clone which is shown in Example 3 to be necessary for the development of the infectious clone.

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Since GBV-B is the virus most closely related to HCV, the present invention also relates to experimental infection of tamarins with the infectious GBV-B clone of the invention or with mutants of the infectious GBV clone to study indirectly the molecular properties of HCV or as a preliminary screen to identify agents which have antiviral activity against HCV. For example, since the predicted internal ribosome entry site (IRES) structure in the 5'UTR of GBV-B is similar to that of HCV (Lemon 1997), the NS3 serine proteases of GBV-B and HCV have been shown to share substrate specificity in vitro (Scarselli 1997), and the 3'UTRs of HCV (Yanaqi 1999) and GBV-B (see Examples) have been shown to be critical for viral infectivity, mutagenesis of these regions in the GBV-B infectious clone may be undertaken to examine IRES function, NS3 serine protease activity or the role of the 3'UTR in viral infectivity in vivo. Where such "mutations" are introduced into the GBV-B clone of the invention to create a "mutated" GBV-B sequence, the mutations include, but are not limited to, point mutations, deletions and insertions. Of course. one of ordinary skill in the art would recognize that the size of the insertions would be limited by the ability of the resultant nucleic acid sequence to be

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properly packaged within the virion. Such mutations could be produced by techniques known to those of skill in the art such as site-directed mutagenesis, fusion PCR, and restriction digestion followed by religation.

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Alternatively, given the significant structural homology that exists between the genomes of GBV and HCV, the infectious GBV-B clone may be used to screen for inhibitors of IRES function or viral enzyme activity (for example, NS3 helicase, NS3 protease, NS2-NS3 protease or NS5B RNA polymerase activity). Such inhibitors may be useful as antiviral agents to HCV since viral enzyme activity and IRES function are known to be critical for HCV replication.

The effect of such inhibitors on the IRES function or viral activity of the GBV-B encoded by the infectious sequence of the invention may be measured by assays known to those of skill in the art to measure directly or indirectly viral replication or viral pathogenicity. Such assays include, but are not limited to, the measurement of virus titer in serum or liver of an infected tamarin by PCR or the measurement of GBV-B viral protein expression in liver cells of an infected tamarin by immunoflourescence or Western blot. course, it is understood that a comparison of results obtained for control tamarins (treated only with infectious nucleic acid sequence) with those obtained for treated tamarins (nucleic acid sequence and antiviral agent) would indicate, the degree, if any, of antiviral activity of the candidate antiviral agent. Of course, one of ordinary skill in the art would readily understand that the tamarins can be treated with the candidate antiviral agent either before or after

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exposure to the infectious nucleic acid sequence of the present invention.

In yet another embodiment, the invention relates to "chimeric nucleic acid sequences" which consist of portions of the infectious nucleic acid sequence of GBV-B and portions of nucleic acid sequences of viruses which are related to GBV-B such as HCV, GBV-C and other members of the Flaviviridae family which do not infect tamarins. In a preferred embodiment, chimeric nucleic acid sequences consist of portions of the infectious nucleic acid sequence of GBV-B and portions of nucleic acid sequences of hepatitis C viruses (HCV) of various genotypes or subtypes; preferably portions of nucleic acid sequence of infectious HCV clones of genotypes la (ATCC accession number PTA-157; Figures 6A-6F), 1b (ATCC accession number 209596; Figures 7A-7F) or 2a (ATCC accession number PTA-153; SEQ ID NO: 4). The nucleic acid sequences taken from GBV-B and HCV can be open-reading frame sequences, and/or sequences from the 5'UTR and/or The gene borders of the HCV genome, including 3'UTR. nucleotide and amino acid locations, have been determined, for example, as depicted in Houghton, M. (1996), and the putative gene borders of the GBV-B are shown in Table 1.

of course, it is understood that the

production of GBV-B/HCV chimeras could include insertion
of specific genes or regions of the infectious GBV-B
clone into an HCV "genomic backbone" (where the HCV
genomic backbone is preferably an infectious nucleic
acid sequence of HCV genotypes 1a, 1b or 2a described
above) or alternatively, could include insertion of

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specific genes (or portions thereof) or regions of an HCV genome into the GBV-B infectious clone of the invention. Of course, where HCV genes or regions are to be inserted into the GBV-B infectious clone, it is to be understood that the inserted HCV sequences may be unmodified or may be mutated in order to examine the effect of the mutation(s) on the function of the inserted HCV gene or region in the chimeric GBV-B-HCV virus.

Such chimeras can readily be produced by methods known to those of ordinary skill in the art.

In one embodiment, GBV-B/HCV chimeras may be made in which 5' or 3' UTR sequences of the GBV-B infectious clone are replaced with the corresponding sequence from an HCV clone. For example, chimeras may be constructed in which the IRES sequence of the infectious GBV-B clone is replaced by the IRES sequence of HCV. Such chimeras can be used in identifying inhibitors of IRES activity which would be useful as antiviral agents, or could be used to examine HCV IRES function in vivo. Alternatively, mutations could be introduced into the HCV IRES contained in the GBV-B clone in order to examine the effect of the mutation(s) on IRES function in vivo.

Alternatively, GBV-B/HCV chimeras may be made in which the 3'UTR sequence of GBV-B is replaced by the 3'UTR sequence of HCV. As the 3' terminal stem-loop structure is believed to be important for initiation of RNA replication and has been shown to be critical for infectivity of HCV <u>in vivo</u>, such chimeras may be used for more detailed analysis of the function of the 3' UTR

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sequence of HCV <u>in vivo</u> and for the testing of candidate antiviral agents.

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In another embodiment, GBV-B/HCV chimeras may be constructed in which the structural or non-structural regions of GBV-B are replaced by corresponding regions of HCV. Such chimeras would be useful in identifying whether the inability of HCV to infect tamarins is due to the inability of HCV's structural region to bind the receptor necessary for infection of tamarins or to the absence of sequences in HCV's nonstructural regions which are necessary for replication in tamarins. For example, the ability to infect tamarins with GBV-B/HCV chimeras in which the non-structural region of GBV-B is replaced by the non-structural region of HCV would indicate that the structural genes of GBV-B are necessary for viral infection in tamarins, and that the inability of HCV to infect tamarins is likely due to its lack of receptors for HCV.

Alternatively, the ability to infect tamarins with GBV-B/HCV chimeras in which the structural region of GBV-B is replaced by the structural region of HCV would indicate that the non-structural genes of GBV-B are critical for viral infection in tamarins, and that the inability of HCV to infect tamarins is likely due to HCV's lack of nonstructural sequences which are necessary for replication in tamarins.

Of course, GBV-B-HCV chimeras may be constructed in which only a portion of the non-structural or structural regions of GBV-B are replaced by the corresponding portions of HCV sequences. For example, a chimera in which only one or two of the three structural genes (C, E1 and E2) of GBV-B are replaced by

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the corresponding HCV structural genes may be made. In one embodiment, nucleic acid sequences comprising the E1 and E2 genes of GBV-B may be replaced by the sequences comprising the HCV E1 and E2 genes. In another embodiment, nucleic acid sequence comprising either the E1 or E2 gene of GBV-B is replaced by sequence encoding either the HCV E1 or E2 gene.

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Alternatively, only a fragment of a GBV-B structural gene in the infectious GBV clone may be replaced with the corresponding HCV gene fragments. For example, the amino terminal of the GBV-B E1 gene may be replaced by the corresponding portion of an HCV E1 gene or an amino terminal portion of the GBV-B E2 gene may be replaced by an amino terminal portion of HCV E2 gene tht containing the HVR1 region. As the structural genes of HCV are believed to be important for neutralization, chimeras containing an HCV structural gene(s) or fragment(s) thereof can be used to develop vaccines against HCV.

In yet another embodiment, chimeras in which individual non-structural genes of GBV-B, such as NS3 RNA helicase, NS3 protease, or the NS5B RNA-dependent RNA polymerase are replaced by the corresponding non-structural genes of HCV may be constructed. Such chimeras would, for example, be useful in identifying inhibitors of viral enzyme activity which would be useful as antiviral agents. Of course, it is understood that in order to construct chimeras in which the polyprotein cleavage sites of the GBV-B remain intact, it may be desirable to replace only a fragment of a nonstructural gene of GBV-B with the corresponding HCV gene fragment.

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The present invention also relates to polypeptides encoded by the nucleic acid sequences of the invention or fragments thereof. In one embodiment, said polypeptide or polypeptides may be fully or partially purified from viruses produced by cells transfected with the nucleic acid sequences of the invention. In another embodiment, the polypeptide or polypeptides may be produced recombinantly from a fragment of the nucleic acid sequences of the invention. In yet another embodiment, the polypeptides may be chemically synthesized.

The present invention further relates to the <u>in vitro</u> and <u>in vivo</u> production of GBV-B, mutated GBV-B or chimeric GBV-B/HCV viruses from the nucleic acid sequences of the invention.

In one embodiment, the sequences of the invention can be inserted into an expression vector that functions in eukaryotic cells. Eukaryotic expression vectors suitable for producing high efficiency gene transfer in vivo are well known to those of ordinary skill in the art and include, but are not limited to, plasmids, vaccinia viruses, retroviruses, adenoviruses and adeno-associated viruses.

In another embodiment, the sequences contained in the recombinant expression vector can be transcribed in vitro by methods known to those of ordinary skill in the art in order to produce RNA transcripts which encode the GBV-B of the invention. The GBV-B of the invention may then be produced by transfecting cells by methods known to those of ordinary skill in the art with either the in vitro transcription mixture containing the RNA

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transcripts or with the recombinant expression vectors containing the nucleic acid sequences described herein.

In assaying the ability of the mutated GBV-B sequences or of the chimeric sequences of the invention to infect tamarins, the virulence phenotype of the virus produced by transfection of tamarins with the sequences of the invention can be monitored by methods known in the art such as measurement of liver enzyme levels (alanine aminotransferase (ALT) or isocitrate dehydrogenase (ICD)) or by histopathology of liver biopsies.

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The present invention also relates to the use of the infectious GBV-B sequence, the mutated GBV-B nucleic acid sequences or the chimeric sequences of the invention to identify cell lines capable of supporting the replication of GBV-B or the chimeras of the invention.

Transfection of tissue culture cells with the nucleic acid sequences of the invention may be done by methods of transfection known in the art such as electroporation, precipitation with DEAE-Dextran or calcium phosphate, or incorporation into liposomes.

In one such embodiment, the method comprises the growing of animal cells <u>in vitro</u> and transfecting the cells with the nucleic acid of the invention, then determining if the cells show indicia of GBV-B or HCV infection. Such indicia include the detection of viral antigens in the cell, for example, by immunofluorescence procedures well known in the art; the detection of viral polypeptides by Western blotting using antibodies specific therefor; and the detection of newly transcribed viral RNA within the cells via methods such

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as RT-PCR. The presence of live, infectious virus particles following such tests may also be shown by injection of cell culture medium or cell lysates into healthy, susceptible animals, with subsequent exhibition of the signs and symptoms of GBV-B infection.

Suitable cells or cell lines for culturing GBV-B or the chimeric GBV-B-HCV include, but are not limited to, lymphocyte and hepatocyte cell lines known in the art.

Alternatively, primary hepatocytes can be cultured, and then infected; or, the hepatocyte cultures could be derived from the livers of infected tamarins. In addition, various immortalization methods known to those of ordinary skill in the art can be used to obtain cell-lines derived from hepatocyte cultures. For example, primary hepatocyte cultures may be fused to a variety of cells to maintain stability.

The invention also provides that the nucleic acid sequences and viruses of the invention be supplied in the form of a kit, alone or in the form of a pharmaceutical composition.

All scientific publication and/or patents cited herein are specifically incorporated by reference. The following examples illustrate various aspects of the invention but are in no way intended to limit the scope thereof.

EXAMPLES

Materials and Methods

Source of GB virus B

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Two tamarin pools VR-806, (American Type Culture Collection) and H205, were used for experimental

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transmission of the GB virus agents to tamarins species

Saguinus mystax and Saguinus oedipus.

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Amplification, cloning and sequence analysis of GBV-B

Viral RNA was extracted from aliquots of the

GB 2/94 serum pool or CT 11/91 liver homogenate with the

TRIZOL system (GIBCO/BRL). Primers used in cDNA

synthesis and PCR amplification were based on the

genomic sequence of GBV-B published by Simons et al

(Simons 1995) shown in SEQ ID NO:3. Long RT-PCR was

performed using Superscript II reverse transcriptase

(GIBCO/BRL) and the Advantage cDNA polymerase mix

(Clontech) as described previously (Tellier 1996). Four

subgenomic regions of GBV-B covering the entire

published sequence (Simons 1995) were amplified from

serum and the PCR products were purified and cloned into

pGEM-9Zf(-) (Promega) or pCR2.1 vector (Invitrogen)

using standard procedures.

The 5' terminus of GBV-B was amplified from serum by using the rapid amplification of cDNA ends (RACE) with dC or dA tailing (GIBCO/BRL) and GBV-B specific antisense primers. Two different approaches were used to determine the 3' terminal sequence of GBV-B. In one approach, GBV-B RNA extracted from serum was circularized with T4 RNA ligase (Promega) and the 5'-to-3'-end-ligated viral RNA was amplified in RT-PCR using specific GBV-B primers. In the second approach, the 5' end of the negative strand GBV-B RNA extracted from the liver homogenate was amplified using the 5' RACE with dC tailing and GBV-B specific sense primers. The PCR products were cloned directly into pCR2.1-TOPO by using the TOPO TA Cloning Kit (Invitrogen).

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The consensus sequence of GBV-B was determined by direct sequencing of PCR products (nucleotides 1-9078 and nucleotides 9130-9359) and by sequence analysis of the clones (nucleotides 1-7135 and nucleotides 7151-9399). Nucleotide positions correspond to those of the infectious clone (pGBB). Analyses of genomic sequences were performed with GeneWorks (Oxford Molecular Group) (Bukh 1995). To determine whether the GenBank data base contained sequences with homology to the GBV-B 3' UTR sequence identified in the present invention, a "Blast" search was performed. The predicted secondary structure of the GBV-B and HCV 3' UTR sequences were determined by the program "mfold" (Genetics Computer Group).

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15 Construction of consensus cDNA clones of GBV-B First, clone pGBB5-1, a consensus clone of GBV-B 2/94 containing the 3' terminus of GBV-B as published by Simons et al was constructed (Simons 1995a). The core sequence of the T7 promoter, a 5' 20 quanosine residue and the sequence of GBV-B (9139 nucleotides) were cloned into pGEM-9Zf(-) vector using NotI/SacI sites. A BamHI site was included at the GBV-B 3' terminus. Digested fragments containing the 25 consensus sequence were purified from subclones and ligated using convenient sites. Next, a second consensus clone of GBV-B, clone pGBB, was constructed by inserting the additional 3' terminal sequence, amplified by PCR from one of the clones obtained by the RACE 30 procedure described above, into pGBB5-1 using XmaI (at position 9114) and BamHI sites. A XhoI site was inserted following the GBV-B 3' terminus. DH5-alpha competent cells (GIBCO BRL) were transformed and 35

selected on LB agar plates containing 100 µg/ml

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ampicillin (SIGMA) and amplified in LB liquid cultures at 30°C for 18-20 hrs (Yanagi 1997). Each cDNA clone was re-transformed to select a single clone, and large-scale preparation of plasmid DNA was performed with a QIAGEN plasmid Maxi kit as described previously (Yanagi 1997). Each clone was genetically stable since the digestion pattern was as expected following retransformation and the complete sequence was the expected one.

Intrahepatic transfection of tamarins with transcribed GBV-B RNA

In 100 μ l reactions, RNA was transcribed in vitro with T7 RNA polymerase (Promega) from 10 μg of linearized template plasmid. The plasmid pGBB5-1 was linearized with BamHI (Promega) and the plasmid pGBB was linearized with XhoI (Promega). The integrity of the RNA was checked by electrophoresis through agarose gel stained with ethidium bromide. Each transcription mixture was diluted with 400 μ l of ice-cold phosphate-buffered saline without calcium or magnesium (SIGMA) and then immediately frozen on dry ice and stored at -80°C. Within 24 hours of synthesis, two transcription mixtures were injected into each tamarin by percutaneous intrahepatic injection guided by ultrasound (Emerson, 1992; Yanaqi 1998, 1999). If the tamarin did not become infected, the same transfection was repeated once. All transfected animals were negative for GBV-A_{SM} as determined by the protocol described previously (Bukh 1997a).

Monitoring of experimental course in tamarins

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Serum samples were collected weekly from the tamarins and monitored for liver enzyme levels [alanine aminotransferase (ALT), gamma-glutamyltranspeptidase (GGT), and isocitrate dehydrogenase (ICD)] by standard methods and for GBV-B RNA by a specific reverse 5 transcriptase-polymerase chain reaction (RT-PCR) assay. Total RNA was extracted from 100 µl of serum using the TRIzol reagent. The RNA pellet was resuspended in 10 mM dithiothreitol (DTT) containing 5% (vol/vol) of RNasin 10 $(20-40 \text{ u/}\mu\text{l})$ (Promega). The RT-nested PCR was performed with primers from the 5' UTR of GBV-B (external primer pair: 5'-CCT AGC AGG GCG TGG GGG ATT TCC-3' and 5'-AGG TCT GCG TCC TTG GTA GTG ACC-3'; internal primer pair: 15 5'-GGA TTT CCC CTG CCC GTC TG-3' and 5'-CCC CGG TCT TCC CTA CAG TG-3'). The reverse transcription was performed with avian myeloblastosis virus reverse transcriptase (Promega) and the external anti-sense primer and nested PCR was performed with AmpliTag DNA polymerase or 20 AmpliTaq Gold DNA polymerase (Perkin Elmer) as described previously (Bukh 1998a). Specificity was confirmed by sequence analysis of selected DNA products. Each set of experiments included a positive control sample (a 10⁻⁶ 25 dilution of GB 8/93, estimated titer 100 genome equivalent (GE)) and appropriate negative control samples. The genome equivalent (GE) titer of GBV-B in positive samples was determined by RT-nested PCR on 10-fold serial dilutions of the extracted RNA (Bukh 30 1998a). One GE was defined as the number of GBV-B genomes present in the highest dilution positive in RTnested PCR. The sensitivity of this RT-nested PCR assay for GBV-B was equivalent to that of our RT-nested PCR 35 assay for HCV (Bukh 1998b), for example, conserved NS3

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primers which had the same sensitivity for GBV-B as the 5' UTR primers could detect HCV at optimal sensitivity in samples with known HCV genome titer. Testing for GBV-A and GBV-A variants was performed by RT-nested PCR assays as described previously (Bukh 1997a).

The consensus sequence of the complete ORF was determined by direct sequencing of overlapping PCR products obtained by long RT-nested PCR on serum from one of the tamarins infected with RNA transcripts as previously described (Yanagi 1997).

Example 1

Transmission of GB Agent in Tamarins

To generate virus pools of the GB agent, tamarins were inoculated intravenously with pooled sera of the eleventh tamarin passage of this agent (Fig. 1). Acute phase sera from a S. mystax tamarin which developed hepatitis were pooled (GB 8/93) and inoculated into additional S. mystax tamarins to generate a second pool of acute phase serum (GB 2/94). Both serum pools contained approximately 10⁸ GE/ml of GBV-B and GBV-A. A 10% liver homogenate (CT 11/91) was prepared from a S. oedipus tamarin which developed hepatitis following inoculation with the twelfth passage of the GB agent. The titer of GBV-B in the liver homogenate was approximately 10⁷ GE/ml. The GB 2/94 serum and CT 11/91 liver samples were used as GBV-B cloning sources in the present study.

Inoculation of eight S. mystax tamarins with ten-fold serial dilutions of the GB 2/94 pool demonstrated that its infectivity titer of GBV-B was 10^8 tamarin 50% infectious doses (TID₅₀) (Fig. 2). The five

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GBV-B infected tamarins all developed acute resolving hepatitis characterized by early appearance of viremia (weeks 1 or 2 p.i.), peak viral titers of 10⁷-10⁸ GE/ml and clearance of viremia after 9-16 weeks (Fig. 2). Two of these tamarins (S. mystax 769 and 777) were infected only with GBV-B and were negative for GBV-A and GBV-Asm, whereas the other three tamarins were infected with both GBV-B and GBV-A_{SM}. A S. mystax tamarin inoculated with the liver homogenate also developed acute resolving hepatitis with peak GBV-B titers of 107 GE/ml and clearance of viremia after 11 weeks. Likewise, four S. mystax tamarins inoculated with dilutions of the GB 8/93 pool developed acute resolving hepatitis with clearance of the GBV-B virus after 11-26 weeks. Thus, GBV-B infection in S. mystax tamarins is characterized by acute hepatitis, early appearance of viremia, high peak viral titers and viral clearance.

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<u>Example 2</u>

Novel 3' Terminal Sequence of GBV-B

The consensus sequence of the complete 5' UTR of GBV-B (nucleotides 1-445) was deduced from 13 clones containing nucleotides 1-283 and 3 clones containing nucleotides 31-445. In addition, the entire 5' UTR sequence was determined by direct sequencing of the amplicons. The sequences of the various clones were highly conserved and the consensus 5' UTR sequence of GBV-B from this pool was identical to that of the previously published sequence for GBV-B (Simons 1995a). It is noteworthy that 13 of 15 clones analyzed from the rapid amplification of cDNA ends (RACE) procedure contained the published GBV-B 5' terminus (A residue)

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and that the same 5' terminus was obtained whether the 5' RACE was performed with dC or dA tailing.

The consensus sequence of the ORF (nucleotides 446-9037) was determined by direct sequencing of PCR products obtained using long RT-PCR (Yanagi 1997). In addition, 3 clones containing nts. 446-7135 (one of these clones had a deletion of nts. 3036-3636), 2 clones containing nts. 2019-3373, 5 clones containing nts. 7151-8261 and 7 clones containing nts. 7521-9037 were analyzed. The sequences of GBV-B clones in this pool were very homogeneous. Evidence of micro-heterogeneity was found at only 70 (0.8%) nucleotide and 36 (1.3%) amino acid positions, scattered throughout the ORF. proportion of amino acid positions with heterogeneity ranged from 0.5-3.2% in different putative gene regions (lowest in NS3 and NS5B; highest in E2 and NS2). GBV-B ORF sequence differed from the published sequence of GBV-B (Simons 1995) at 34 (0.4%) nucleotide and 12 (0.4%) deduced amino acid positions, respectively (Table

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Table 1

Nucleotide and amino acid differences among GBV-B (Simons 1995a), the consensus sequence of GBV-B recovered from a virus pool used as the cloning source (GBV-B, 2/94) and the infectious clone of GBV-B (pGBB).

5	Genomic Region*	Position nt [aa]	Nucleotide			Amino Acid		
				GBV-B			GBV-B	· ·-·
			<u>GBV-B</u>	2/94	pGBB	GBV-B	2/94	<u>PGBB</u>
	5' UTR (1-445)							
	C (446-913)		-	_	_			
	E1 (914-1489)	1030	C	T	T			
	E2 (1490-2641)	1498	T	C (t)	C			_
		1628 [395]	G	A (g)	A	V	I (V)	I
• •		2552 [703]	G C N	A (g)	A	D	N (D)	N
10		2562,2563 [706]	C,A	A,C	A,C	P	Н	Н
		2566	т	т	T			
		2625 [727]	Ċ	T	T	A	v	v
	NS2 (2642-3385)	2647	c	T (c)	T	A	V	v
	1 102 (2012 3303)	2816 [791]	C	T	T	L	F	F
		2855 [804]	Ä	Ğ	Ġ	T	A	A
		3235	A	G	G	•		r.
	NS3 (3386-5125)	3475**	c	C (t)	T			
		3760	Ċ	T (c)	T			
15		4114	Ċ	T	T			
15		4117	C	A	Ā			
		4177	T	С	c			
		4615	C	T	T			
	NS4A (5126-5290)							
	NS4B (5291-6034)	5329	C	T	T			
		5332	T	С	С			
		5350	A	С	C			
		5455	C	T (c)	T			
	NS5A (6035-7267)	6413	T	A (t)	A	L	M (L)	M
20		[1990]						
20		6577	G	T	T			
		6690	T	C (t)	C	I	T(I)	T
		[2082]						
		6965	T	C (t)	C	S	P (S)	P
		[2174]	_		_			
		7015	A	G (a)	G	~	_	_
		7128	G	A	A	G	E	E
		[2228] 7138**	A	A	G			
		7142	A	G	G	т	A	A
25		[2233]	A	G	G	1	А	A
	NS5B (7268-9037)	7282	Т	C (t)	С			
	10000 (7200 0007)	7849	Ċ	A	A			
		7852	C	T	T			
		8942	Ğ	A (g)	A	v	I (V)	I
		[2981]	_	13/		-	_ \ \ \ /	-
		8971	T	С	С			
		9026	c	T (c)	T			
	3' UTR (9038-	9067	T	C	C			
30	9399)							
30		Poly(U)	27 nts	11-23 nts	23 nts			
		9134	Deletion	С	С			
		9141-9399	ND	259 nts	259 nts			

^{*}Nucleotide positions corresponding to pGBB. Putative gene borders defined as suggested by homology with HCV (Muerhoff 1995). No homology was observed at the NS2-NS3 junction.

^{**}Positions that differ between the cloning source (GBV-B 2/94) and the infectious clone of GBV-B (pGBB). The change introduced into pGBB at position 7138 introduced an artificial SalI site. nd: Not determined. Nucleotides and amino acids shown in parenthesis were found as a minor species in the cloning source (GBV-B, 2/94

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The sequence for the 3' UTR is shown in Figure 3. Additional 3' UTR sequence was initially identified by performing RT-PCR across 5'-to-3'-end-ligated viral RNA extracted from serum. In all 4 clones with GBV-B sequences, the 5' UTR was truncated compared to the 5 published sequence (simon 1995a). However, whereas one clone (29c) had the exact 3' terminus previously published by Simons et al. (Simons 1995a), the three other clones (29a, 29b, 29d) had 150 additional terminal 10 nucleotides. Compared with the published sequence, all four clones had a single nucleotide insertion (C residue) at position 9134. Next, RACE using dC-tailing only was performed on the 5' end of the negative-strand RNA extracted from the liver homogenate. All 11 clones 15 analyzed had additional sequences at the 3' terminus. Compared with the published GBV-B sequence, two clones (qb6, qb23) had 259 additional nucleotides, 8 clones (gb9, gb19, gb20, gb21, gb24, gb25, gb30, gb35) had 236 20 additional nucleotides and 1 clone (qb8) had 232 additional nucleotides. Moreover, all of these clones had the insertion at position 9134. The 3' UTR sequences among the various clones were highly conserved 25 (Fig. 3). To demonstrate that the terminal 22 nucleotides found only in clones qb6 and qb23 existed in circulating viruses, RT-nested PCR was performed on 10fold serially diluted RNA extracted from the serum pool GB 2/94 using an RT and external antisense primer 30 deduced from this sequence. GBV-B RNA was detected at a dilution of 10⁻⁷ and the sequence of the amplicon was identical to the sequence recovered from the liver homogenate. Thus, the 3' UTR of GBV-B consists of a 35 short sequence of 30 nucleotides followed by a 11-24

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nucleotide-long poly (U) tract (single C residues were observed in GBV-B from the liver homogenate) and a 3' terminal sequence of at least 309 nucleotides. The new GBV-B 3' UTR sequence did not have significant homology to any of the sequences deposited in the GenBank database. A prediction of the secondary structure of the 3' UTR sequence is shown in Figure 4. The most notable feature of the secondary structure is a highly stable stem-loop structure at the very 3' end consisting of 47 nucleotides.

Example 3

The pGBB Clone of GBV-B is Infectious in vivo

The infectivity of RNA transcripts from the consensus clone pGBB5-1 which encompassed only the published GBV-B sequence (Simons 1995) was first tested. Within the GBV-B sequence there were no deduced amino acid differences and only 2 nucleotide differences (at nucleotide positions 3475 and 7138) between the consensus sequence of the cloning source (GBV-B 2/94) and the sequence of pGBB5-1 clone. In addition, the 3' UTR of pGBB5-1 had a deletion at nucleotide position 9134 and was missing the 3' terminal 259 nucleotides (Fig. 3). Prior to transcription, the pGBB5-1 clone was linearized at the BamHI site with digestion at the exact GBV-B 3' terminus. The RNA transcripts from pGGB5-1 were injected into the liver of two tamarins (S. mystax 797 and 815). GBV-B RNA was not detected in weekly serum samples collected during 17 weeks of follow-up. As the susceptibility of these two tamarins to GBV-B was subsequently demonstrated by experimental infection using a GBV-B virus pool, the consensus clone pGBB5-1

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which lacks the 3' terminal sequence of GBV-B is thus not infectious in vivo.

Next, the infectivity of RNA transcripts from the full-length consensus GBV-B cDNA clone pGBB was The pGBB clone was identical to the pGBB5-1 clone except in the 3' UTR. Thus, in addition to a 5' UTR of 445 nucleotides, an ORF of 8592 nucleotides encoding 2864 amino acids and a 3' UTR of 103 nucleotides, the pGBB clone also contains an additional 259 nucleotides in its 3' UTR. pGBB was linearized at the XhoI site which added an additional C residue at the 3' end of the transcribed GBV-B RNA. When RNA transcripts from the pGBB clone were injected into the liver of two tamarins (S. mystax 816 and 817), both tamarins became infected with GBV-B with viremia at week 1 p.i. and peak viral titers of 108 GE/ml (Fig. 5). consensus sequence of PCR products of the complete ORF, amplified from serum obtained during week 2 p.i. from one tamarin (S. mystax 817), was identical to the sequence of pGBB, including at the two positions which differed from the consensus sequence of the cloning source and from the published sequence of GBV-B (Table By performing RT-PCR as desired above, it was demonstrated that the very 3' terminal GBV-B sequence of pGBB existed in the circulating viruses in this tamarin. Within two weeks of the transfection both tamarins developed hepatitis with dramatically elevated liver enzyme levels (Fig. 5). Thus, the pGBB clone is infectious in vivo whereas the clone pGBB5-1 which lacks the last 259 nucleotides was not.

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• WHAT IS CLAIMED IS:

- 1. An isolated nucleic acid molecule which encodes GB virus-B, said molecule capable of expressing said virus when transfected into cells.
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 2. The nucleic acid molecule of claim 1,
 wherein said molecule encodes the amino acid sequence of
 SEQ ID NO:2.
- 3. The nucleic acid molecule of claim 2, wherein said molecule comprises the nucleic acid sequence of SEQ ID NO:1.
 - 4. A DNA construct comprising a nucleic acid molecule according to claim 1.
- 5. A DNA construct comprising a nucleic acid molecule according to claim 3.
 - 6. An RNA transcript of the DNA construct of claims 4 or 5.
- 7. A cell transfected with the DNA construct of claims 4 or 5.
 - 8. A cell transfected with RNA transcripts of claim 6.
- 9. A GB virus-B polypeptide produced by the cell of claim 7.
 - $$\tt 10.$\ A$ GB virus-B polypeptide produced by the cell of claim 8.
- 11. A GB virus-B produced by the cell of claim 7.
- $$\tt 12.$$ A GB virus-B produced by the cell of claim 8.

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13. A GB virus-B whose genome comprises a nucleic acid molecule according to claim 1.

- 14. A GB virus-B whose genome comprises a nucleic acid molecule according to claim 3.
- 15. A method for producing a GB virus-B comprising transfecting a host cell with the DNA construct of claims 4 or 5.

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- 16. A method for producing a GB virus-B comprising transfecting a host cell with the RNA transcript of claim 6.
 - 17. A composition comprising a nucleic acid molecule of claim 1 suspended in a suitable amount of a pharmaceutically acceptable diluent or excipient.
 - 18. A composition comprising a nucleic acid molecule of claim 3 suspended in a suitable amount of a pharmaceutically acceptable diluent or excipient.
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 19. A nucleic acid molecule comprising a chimeric virus genome, said genome being a GB virus-B genome according to claim 1 in which a 3' or 5' UTR sequence of the genome is replaced by a corresponding region of the 3' or 5' UTR sequence of a hepatitis C virus genome.
 - 20. The nucleic acid molecule of claim 19, wherein a 3' UTR sequence of the genome of a GB virus-B is replaced by a corresponding 3' UTR sequence of a hepatitis C virus genome.
 - 21. The nucleic acid molecule of claim 20, wherein the 3' UTR sequence is the 3' UTR terminal stem loop sequence.

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22. The nucleic acid molecule of claim 19, wherein a 5' UTR sequence of the genome of a GB virus-B has been replaced by a corresponding 5' UTR sequence of a hepatitis C virus genome.

5 23. The nucleic acid molecule of claim 22, wherein the 5' UTR sequence is the IRES sequence.

- 24. A nucleic acid molecule comprising a chimeric virus genome, said genome being a GB virus-B genome according to claim 1 in which the non-structural region of the genome of a GB virus-B has been replaced by the non-structural region of a hepatitis C virus genome.
- 25. The nucleic acid molecule of claim 24, wherein at least one gene from the non-structural region of the genome of a GB virus-B has been replaced by the corresponding gene from the non-structural region of a hepatitis C virus genome.

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26. The nucleic acid molecule of claim 25, wherein the gene from the non-structural region is selected from the group consisting of NS3 protease, NS3 RNA helicase, or NS5B RNA polymerase.

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- 27. A nucleic acid molecule comprising a chimeric virus genome, said genome being a GB virus-B genome according to claim 1 in which the structural region of the genome of a GB virus-B has been replaced by the structural region of a hepatitis C virus genome.
- 28. The nucleic acid molecule of claim 27, wherein at least one gene from the structural region of the genome of a GB virus-B has been replaced by the

- 35 -

corresponding gene from the structural region of a hepatitis C virus genome.

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- 29. The nucleic acid molecule of claim 28, wherein the gene from the structural region is selected from the group consisting of E1, E2 or C.
- 30. The nucleic acid molecule of claim 28, wherein the E1 and E2 genes from the structural region of the genome of a GB virus-B have been replaced by the E1 and E2 genes of a hepatitis C virus genome.
- 31. The nucleic acid molecule of claim 28, wherein the E1 gene from the structural region of the genome of a GB virus-B has been replaced by the E1 gene of a hepatitis C virus genome.
- 32. The nucleic acid molecule of claim 28, wherein the E2 gene from the structural regions of the genome of a GB virus-B has been replaced by the E2 gene of a hepatitis C virus genome.
- 33. A DNA construct comprising the nucleic acid molecule of claims 19, 24 or 27.
- 34. An RNA transcript of the DNA construct of claim 33.
- 35. A virus whose genome comprises a nucleic acid molecule according to claims 19, 24 or 27.
- 36. A nucleic acid molecule comprising a chimeric virus genome, said genome being a hepatitis C virus genome in which a 3' or 5' UTR sequence of the genome is replaced by a corresponding region of the 3' or 5' UTR sequence of a GB virus-B genome according to claim 1.

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37. A nucleic acid molecule comprising a chimeric virus genome, said genome being a hepatitis C virus genome in which the non-structural region of the

genome has been replaced by the non-structural region of a GB virus-B genome according to claim 1.

38. A nucleic acid molecule comprising a chimeric virus genome, said genome being a hepatitis C virus genome in which the structural region of the genome has been replaced by the structural region of a GB virus-B genome according to claim 1.

39. A polypeptide encoded by the nucleic acid molecule of claims 19, 24 or 27.

15 40. A polypeptide encoded by the nucleic acid molecule of claims 36, 37 or 38.

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FIG. 1

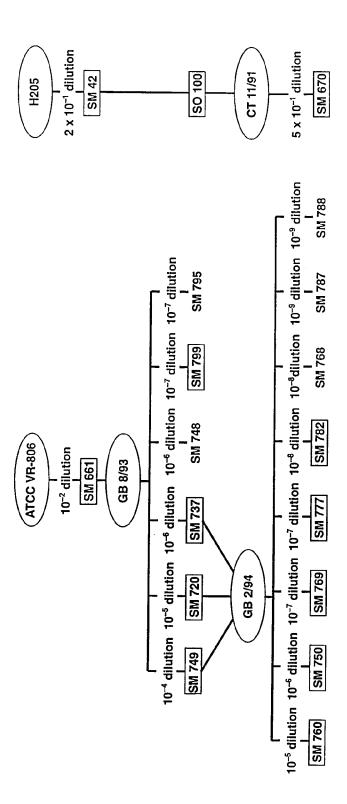
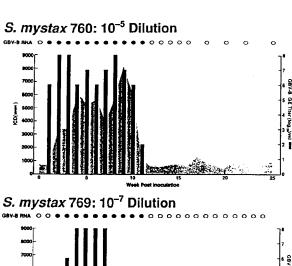
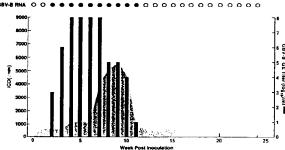
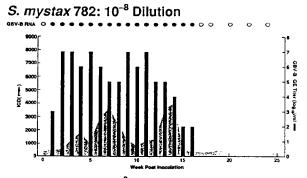
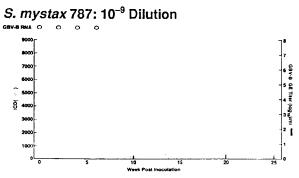


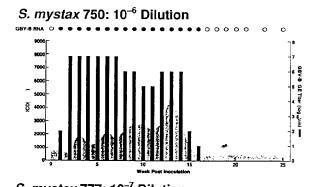
FIG. 2

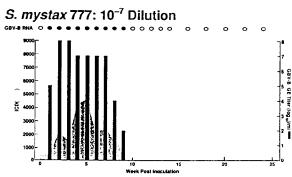


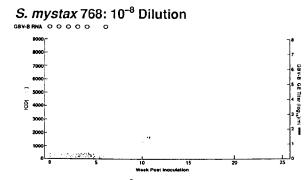












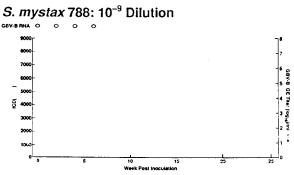


FIG. 3

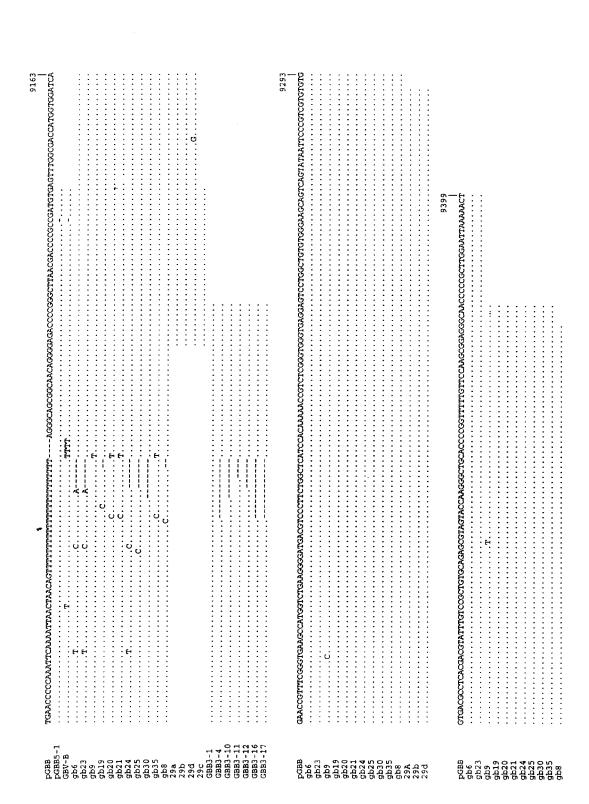
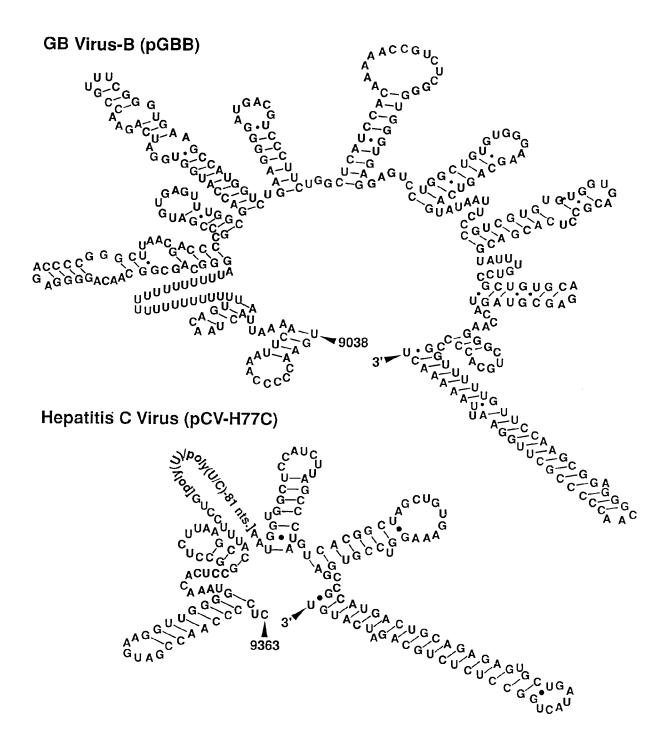


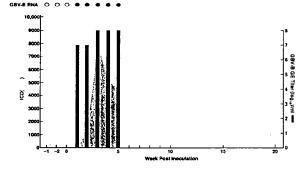
FIG. 4



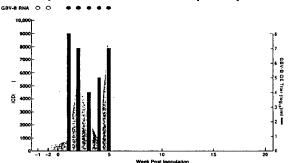
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FIG. 5





S. mystax 817: RNA Transcripts of pGBB



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GCAATGAGGG TIV	300GGTGG	GCGGGGATGGC	TOCIGICICC	CCGIGGCICT	650
CGGCCTAGCT GG	3GCCCAC	AGACCCCCGG	CGIAGGICGC	GCAATTIGGG	700
TAAGGICATC GA	IACCCTIA	CGIGCGGCTT	CCCCCACCIC	ATGGGGTACA	750
TACCOCTCGT CC	TOOOGE	CTTGGAGGG	CIGCCAGGGC	CCTGGCGCAT	800
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YHVINDCPNS SIVYEAA				250
CKLPITQLRR HIDLLW	SAT LCSALYVEDI	CGSVFLVGQL	FIFSPRRHWI	300
TODONCSIYP CHITCHE				350
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RCDLEDRDRS ELSPLLI				700
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ILRRHVGPGE	GAVQWMNRLI	AFASRGNHVS	PIHYVPESDA	AARVIAILSS	1950
LIVIQLLRRL	HOWISSECTT	PCSGSWLRDI	WDWICEVLSD	FKIWLKAKIM	2000
PQLPGIPFVS	CORGYRGVWR	GDGIMHIRCH	CCAETICHVK	NGIMRIVGPR	2050
TCRNMWSGIF	PINAYTIGPC	TPLPAPNYKF	ALWRVSAEEY	VEIRRVGDFH	2100
YVSGMIIDNL	KCPCQIPSPE	FFTELDGVRL	HRFAPPCKPL	LREEVSFRVG	2150
LHEYPVGSQL	PCEPEPDVAV	LISMLIDPSH	ITAEAAGRRL	ARGSPPSMAS	2200
SSASQLSAPS	LKATCIANHD	SPDAELIEAN	LLWRQEMGGN	TTRVESENKV	2250
VILDSFDPLV	AEEDEREVSV	PAEILRKSRR	FARALPWAR	PDYNPPLVET	2300
WKKPDYEPPV	VHGCPLPPPR	SPPVPPPRKK	RIVVLTESTL	STALAFLATK	2350
SFGSSSTSGI	TCDNTTTSSE	PAPSGCPPDS	DVESYSSMPP	LEGEPGDPDL	2400
SDGSWSIVSS	GADIEDVVCC	SMSYSWIGAL	VTPCAAEEQK	LPINALSNSL	2450
LRHHNLVYST	TSRSACQRQK	KVIFDRLQVL	DSHYQDVLKE	VKAAASKVKA	2500
NLLSVEFACS	LTPPHSAKSK	FGYGAKDVRC	HARKAVAHIN	SWKDLLEDS	2550
VIPIDITIMA	KNEVFCVQPE	KGGRKPARLI	VFPDLGVRVC	EKMALYDVVS	2600
KLPLAVMGSS	YGFQYSPGQR	VEFLVQAWKS	KKTPMGFSYD	TRCFDSIVIE	2650
SDIRTEEAIY	QCCDLDPQAR	VAIKSLIERL	YVGGPLINSR	GENCGYRRCR	2700
ASGVLTTSCG	NTLTCYTKAR	AACRAAGLQD	CIMLVCGDDL	VVICESAGVQ	2750
EDAASLRAFT	EAMTRYSAPP	${\tt GDPPQPEYDL}$	ELITSCSSNV	SVAHDGAGKR	2800
VYYLTRDPTT	PLARAAWETA	RHTPVNSWLG	NIIMFAPTIW	ARMILMIHFF	2850
SVLIARDQLE	QALNCETYGA	CYSIEPLDLP	PIIQRLHGLS	AFSLHSYSPG	2900
EINRVAACLR	KLGVPPLRAW	RHRARSVRAR	LLSRGGRAAI	CCKYLFIWAV	2950
RTKLKLTPIA	AAGRLDLSGW	FTAGYSGGDI	YHSVSHARPR	WFWFCLLLLA	3000
AGVGIYLLPN	R				3011

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	20	30	40	50	
10	1234567890				
1234567890	TGATGGGGGC	T724201030	TS24201030	עלליקטיסטע	50
	TCTTCACGCA				100
	CCTCCAGGAC				150 150
					200
	GAGTACACCG				250
	CTCAATGCCT				300
	GIGIIGGGIC				350
	GTGCCCCGGG				
	CICAAAGAAA				400
	TTCCCGGGGG				450
	GGGCCCCAGG				500
	AACCICGIGG				550
	AGGGCCTGGG				600
	CCTGGGGTGG				650
	GGGGCCCCAC				700
	GATACCCTTA				750
	CGGCGCCCC				800
GETGTCCGGG	TICIGGAGGA	CGGCGTGAAC	TATGCAACAG	GGAACTTGCC	850
CCGTTCCTCT	TTCTCTATCT	TCCTCTTGGC	TCTGCTGTCC	TGTTTGACCA	900
TCCCAGCTIC	CGCTTATGAA	GIGCGCAACG	TGTCCGGGAT	ATACCATGIC	950
ACGAACGACT	GCTCCAACTC	AAGCATIGIG	TATGAGGCAG	CGGACGIGAT	1000
CATGCATACT	CCCCCCCTTCCCC	TGCCCIGIGI	TCAGGAGGGT	AACAGCTCCC	1050
GITGCTGGGT	AGCGCTCACT	CCCACGCTCG	CGGCCAGGAA	TGCCAGCGIC	1100
CCCACTACGA	CAATACGACG	CCACGICGAC	TTGCTCGTTG	GGACGGCTGC	1150
TITICIGCICC	CCTATGTACG	TGGGGGATCT	CTGCGGATCT	ATTITICCICG	1200
TCTCCCAGCT	GITCACCTIC	TOGOCTOGOC	_GGCATGAGAC	AGIGCAGGAC	1250
	CAATCTATCC				1300
GGATATGATG	ATGAACIGGT	CACCIACAAC	AGCCCTAGIG	GIGICGCAGI	1350
				GGCCCACTGG	1400
GGAGTCCTGG	CGGGCCTTGC	CTACTATICC	ATGGTAGGGA	ACTGGGCTAA	1450
				GAGACCCACA	1500
CGACGGGAG	GCIGCCGGC	CACACCACCI	CCGGGTTCAC	GICCCITITC	1550
				ACGGCAGCTG	1600
				CAAACIGGGT	1650
TCTTTGCCGC	GCIGITITAC	GCACACAAGI	TCAACICGIC	COOGLIGCOCC	1700
GAGCGCATGC	CCAGCTGCCG	CCCCATTGAC	TOGITOGCCC	: AGGGGTGGGG	1750
				CCTTATTCCT	1800
				CAGGIGIGI	1850
				G GGACCACOGA	1900

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10	20	30	40	50	
1234567890		1234567890			
TOGITOCOGI	=	ATAGCTGGGG			1950
		CCGCCACAAG			2000
		CACTAAGACG			2050
		GCACCITGAT			2100
	-	TACACAAAAT			2150
		CTACCCATAC			220 0
		TTAAGGITAG			2250
		TGCAATTGGA			2300
		AGAACTCAGC			2350
		GIGCITICAC			2400
		CAGAACATCG			2450
		CICCITIGCA			2500
011001	- - · ·	CAGACGCGCG			2550
	-	GCTGAGGCCG			2600
		CCCACCCAT			2650
		ACATTAAGGG			2700
CGTATGCTTT	TTATGGCGTA	TGGCCGCTGC	Techecicer	ACIGGCGITA	2750
CCACCACGAG	CTTACGCCTT	GGACCGGGAG		CCICCCCCCCC	2800
	GTAGGTCTGG		· · ·	TACTACAAAG	2850
TGTTTCTCAC	TAGGCICATA	TOGIGGITAC			2900
	TGCAAGIGIG			COCCACCOCC	2950
		CGIGIGCGGI			3000
				' GGIGCICCAG	3050
GCTGGCATAA				GCTCATTCG	3100
TOCATOCATO	= :			GECCAAATGG	3150
				TAACCATCIT	3200
				TICCCGICCC	3250
				AICACCIGGG	3300
				ACCCGICICC	3350
				A GICTOGAAGG	3400
				CAACAAACGC	3450
				GCACAAGAAC	3500
				AATCTTTCCT	3550
				r ggcgctggct	3600
				r Gracaccaat	3650
				G COCCCTCCAT	3700
				C ACGAGACATG	3750
CIGATGICAT	TOCCETIECE	CGGGGAGGG	G ACAGCAGGG	G AAGICIACIC	3800

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10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
TCCCCCAGGC	CCGICICCIA	CCTGAAAGGC	TCCTCCGGGIG	GICCATIGCT	3850
TICCCCTICG	GGGCACGICG	TGGGGGICIT	CCCGCCTCCT	GIGIGCACCC	3900
GGGGGGICGC	GAAGGCGGIG	GACTICATAC	CCGTTGAGIC	TATGGAAACT	3950
ACCATGCGGT	CICCGGICIT	CACAGACAAC	TCAACCCCCC	CCCCICIACC	4000
GCAGACATTC	CAAGIGGCAC	ATCTGCACGC	TCCTACTGGC	AGCGGCAAGA	4050
GCACCAAAGT	GCCGGCTGCG	TATGCAGCCC	AAGGGTACAA	GGIGCICGIC	41.00
CIGAACCCGI	CCGITGCCCC	CACCTTAGGG	TTTGGGGGGT	ATATGICCAA	4150
GGCACACGGT	ATOGACOCTA	ACATCAGAAC	TGGGGTAAGG	ACCATTACCA	- 4200
COGGCCCCTC	CATTACGIAC	TCCACCTATG	GCAAGITCCT	TECCGACCET	4250
GGCTGTTCTG	GGGGGCCTA	TGACATCATA	ATATGTGATG	AGIGCCACIC	4300
AACTGACTCG	ACTACCATCT	TGGGCATCGG	CACAGICCIG	CACCAACCCG	4350
AGACGGCTGG	AGCGCGGCTC	GICGICCICG	CCACCGCTAC	ACCTCCGGGA	4400
TOGGTTACOG	TGCCACACCC	CAATATOGAG	GAAATAGGCC	TGTCCAACAA	4450
TGGAGAGATC	CCCTTCTATG	GCAAAGCCAT	CCCCATTGAG	CCCATCAACG	4500
GGGGGAGGCA	TCTCATTTTC	TGCCATTCCA	AGAAGAAATG	TCACCACCIC	4550
GCCGCAAAGC	TGACAGGCCT	CGGACTGAAC	GCIGIAGCAT	ATTACCGGGG	4600
CCTTGATGIG	TCCGTCATAC	CGCCTATCGG	AGACGICGIT	GICGIGGCAA	4650
CAGACGCTCT	AATGACGGGT	TTCACCGGCG	ATTTTGACTC	AGIGATOGAC	4700
TGCAATACAT	GIGICACCCA	GACAGTOGAC	TICAGCIIGG	ATCCCACCTT	4750
CACCATIGAG	ACGACGACCG	TGCCCCAAGA	cccccicicc	CGCTCGCAAC	4800
GGCGAGGTAG	AACTGGCAGG	CGTACGAGTG	GCATCTACAG	GITIGIGACT	4850
CCAGGAGAAC	GGCCTCGGG	CATGITCGAT	TCTTCCGTCC	TGIGIGAGIG	4900
CTATGACGCG	GGCTGTGCTT	GGTATGAGCT	CACGCCCCCT	GAGACCTCGG	4950
TTAGGTTGCG	GGCTTACCTA	AATACACCAG	GGTTGCCCGT	CTGCCAGGAC	5000
CATCIGGAGT	TCTGGGAGAG	CGTCTTCACA	GGCCTCACCC	ACATAGATEC	5050
CCACTTCCTG	TCCCAGACTA	AACAGGCAGG	AGACAACTTT	CCTTACCTCG	5100
TGGCATATCA	AGCIACAGIG	TGCGCCAGGG	CICAAGCICC	ACCTCCATCG	5150
TGGGACCAAA	TGTGGAAGTG	TCTCATACGG	CIGAAACCIA	CACTGCACGG	5200
GCCAACACCC	CTGCTGTATA	GGCTAGGAGC	CGTCCAAAAT	GAGGTCATCC	5250
TCACACACCC	CATAACTAAA	TACATCATGG	CATGCATGTC	GGCTGACCTG	5300
GAGGTOGTCA	CTAGCACCTG	GGIGCIGGIA	GGCGGAGTCC	TIGCAGCITT	5350
GGCCGCATAC	TGCCTGACGA	CAGGCAGIGI	GGICATIGIG	GGCAGGATCA	5400
TCTTGTCCGG	GAAGCCAGCT	GICGIICCCG	ACAGGGAAGT	CCTCTACCAG	5450
GAGITOGATG	AGATOGAAGA	GIGIGCCICA	CAACTTCCTT	ACATOGAGCA	5500
		AATTCAAGCA			5550
		GAGGCTGCTG			5600
				ATTTCATCAG	5650
CGGAATACAG	TACCTAGCAG	GCTTATCCAC	TCTGCCTGGA	AACCCCCGCGA	5 70 0

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10	20	30	40	50	
1234567890 1	234567890	<u>1234567890</u>	1234567890	1234567890	
TAGCATCATT G	ATGGCATTT	ACAGCTICIA	TCACTAGCCC	GCTCACCACC	5750
CAAAACACCC T	CCIGITIAA	CATCTTGGGG	CGATCCCTCC	CIGCCCAACT	5800
CGCTCCTCCC A	ACCOCTECCET	CAGCITICGI	GGGGGGGGC	ATCGCCCGGAG	5850
CCCCTCTTCC C	AGCATAGGC	CTTGGGAAGG	TECTOSTOGA	CATCTTGGCG	5 90 0
GGCTATGGGG C	AGGGGIAGC	CGGCGCACIC	GIGGCCTTIA	AGGICATGAG	5950
CCCCCACCIG C	CCICCACCG	AGGACCIGGI	CAACITACIC	CCTGCCATCC	6000
TCTCTCCTGG T	GCCCIGGIC	GICGGGGICG	TGTGCGCAGC	AATACTGOGT	6050
CCCCACCICC C	ECCCCGGGAGA	GGGGGCIGIG	CAGIGGAIGA	ACCGGCTGAT	610 0
ACCOPTOCCT T	CGCGGGTA	ACCACGICIC	CCCTACGCAC	TATGIGCCIG	6150
AGAGOGAOGC I	CCACCACGI	GICACICAGA	TOCICICIAG	CCTTACCATC	6200
ACTCAACTGC T	GAAGOGGCT	CCACCAGIGG	ATTAATGAGG	ACIGCICIAC	6250
GCCATGCTCC G	ECTCGTGGC	TAAGGGATGT	TIGGGATIGG	ATATGCACGG	6300
TGITGACTGA C	CTTCAAGACC	TGGCTCCAGT	CCAAACICCT	GCCGCGGTTA	6350
CCCCCGAGTCC C	CTTTCCTGTC	ATGCCAACGC	GGGTACAAGG	CACTCTCCCC	6400
GGGGGACGGC A	ATCATGCAAA	CCACCTGCCC	ATGCGGAGCA	CAGATOGCOG	6450
GACATGICAA A	AAACGGTTCC	ATGAGGATCG	TAGGGCCTAG	AACCTGCAGC	6500
AACACGIGGC A	ACGGAACGIT	CCCCATCAAC	GCATACACCA	CCCCACCTTC	6550
CACACCCTCC C	ACCECCEC	ACTATTCCAG	GGCGCTATGG	CCCCTC	6600
CTGAGGAGIA C	CICCACCIT	ACCCGIGICG	GGGATTTCCA	CTACGTGACG	6650
GGCATGACCA C	TGACAACGT	AAAGTGCCCA	TGCCAGGTTC	CCCCCCCCA	6700
ATTCTTCACG C	EAGGIGGAIG	GAGTGCGGTT	GCACAGGIAC	GCTCCGGCGT	6750
GCAAACCICT I	CTACGGGAG	GACGTCACGT	TOCAGGICGG	GCTCAACCAA	6800
TACTTOGICG C	EGTCGCAGCT	CCCATGCGAG	CCCGAACCGG	ACGIAACAGI	6850
GCTTACTTCC A	ATGCTCACCG	ATCCCTCCCA	CATTACAGCA	GAGACGCCTA	6900
AGOGTAGGCT G	EGCTAGAGGG	TCTCCCCCCT	CTTTAGCCAG	CICAICAGCT	6950
AGCCAGTIGT C	CICCCCCIIC	TTTGAAGGCG	ACATGCACTA	CCCACCATGA	7000
CTCCCCCGGAC G	CIGACCICA	TOGAGGOCAA	CCICITGIGG	CGGCAGGAGA	7050
TGGGCGGAAA C	CATCACTOGC	GIGGAGICAG	AGAATAAGGT	AGIAATICIG	7100
GACTOTTTCG A	AACCGCTTCA	CCCCCACCCC	GATGAGAGGG	AGATATCCGT	7150
CCCCCCCCAG A	ATCCTGCGAA	AATCCAGGAA	GITCCCCTCA	CCGTTCCCCA	7200
TATGGGCACG C	CCCGGACTAC	AATCCTCCAC	TOCTAGAGIC	CIGGAAGGAC	7250
CCGGACTACG I	ICCCICCGGI	GGTACACGGA	TGCCCATTGC	CACCIACCAA	7300
GGCTCCTCCA A	ATACCACCTC	CACGGAGAAA	GAGGACGGII	GICCIGACAG	7350
AATCCAATGT (GICLICIGCC	TTGGCGGAGC	TCGCCACTAA	GACCIICGGI	7400
AGCTCCGGAT (CGICCGCCGT	TGATAGCGGC	ACGGGGACCG	COCTTOCTGA	7450
CCIGGCCICC (GACGACGGIG	ACAAAGGATC	CGACGITGAC	TOGIACICCI	7500
CCATGCCCCC (CCTTGAAGGG	GAGCCGGGGG	ACCCCGATCI	CAGCGACGGG	7550
TCTTGGTCTA (CCGIGAGIGA	GGAGGCTAGT	GAGGATGICC	TCIGCIGCIC	7600

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10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
AATGICCIAT	ACGIGGACAG	CCCCCTGAT	CACGCCATGC	GCTGCGGAGG	7650
		COGTTGAGCA			7700
		ATCCCGCAGC			7750
GGTCACCTTT	CACAGATICC	AAGTOCTGGA	TGATCATTAC	CCCCACCTAC	7800
		GOGTOCACAG			7850
		GACGCCCCA			7900
		TCCGGAACCT			7950
ACATOOGCTC	CGIGIGGGAG	GACTICCICG	AACACACTGA	AACACCAATT	8000
GACACCACCA	TCATGGCAAA	AAGIGAGGIT	TICIGOGICC	AACCAGAGAA	8050
GGGAGGCCGC	AAGCCAGCTC	GCCTTATCGT	ATTOCCAGAC	CIGGGAGITC	8100
GIGIATGCGA	CAACATCCCC	CTTTACGACG	TEGICICCAC	CCTTCCTCAG	8150
GCCGTGATGG	GCTCCTCATA	CGGATTICAA	TACTOCCCA	AGCAGCGGGT	8200
CGAGTTCCTG	GIGAATACCT	GGAAATCAAA	GAAATGCCCT	ATGGGCTTCT	8250
CATATGACAC	CCCCIGITIT	GACTCAACGG	TCACTGAGAG	TGACATTCGT	8300
GTTGAGGAGT	CAATTTACCA	ATGITGIGAC	TTGGCCCCCG	AGGCCAGACA	8350
		AGCGGCTTTA			8400
		GGTTATCGCC			8450
		TACCCTCACA			8 50 0
		TOCAGGACTG			8550
		GAAAGCGCGG			8 60 0
GCCCTACGAG	CCTTCACGGA	GGCTATGACT	ACCIATICCG	CCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	8650
GGATCCGCCC	CAACCAGAAT	ACGACCTGGA	GCTGATAACA	TCATGITCCT	8700
CCAATGIGIC	AGTOGOGCAC	GATGCATCIG	GCAAAAGGGI	ATACIACCIC	8750
		CCTTGCACGG			8800
ACACACTOCA	. ATCAACICIT	'GGCTAGGCAA	TATCATCATC	TATGCGCCCA	8850
				CATCCTICIA	8900
GCTCAAGAGC	AACTTGAAAA	AGCCCIGGAT	TGICAGATCI	ACGGGGCTTG	8950
CTACTCCATT	GAGCCACTIC	ACCIACCICA	GATCATIGA	A CGACTCCATG	9000
GTCTTAGCGC	ATTTACACTO	CACAGITACI	CICCAGGIG	A CATCAATAGG	9050
GTGGCTTCAT	GCCTCAGGAA	ACTTGGGGLA	CCACCCTTGC	GAACCIGGAG	9100
ACATOGGGC	AGAAGIGICC	GOGCTAAGCT	ACTGTCCCAC	GCCCCCACCC	9150
CCGCCACTTC	TOGCAGATAC	CICITIAACI	GGGCAGIAA	GACCAAGCTT	9200
AAACTCACTO	CAATCCCGGC	COCCICCCAC	CTGGACTIG	r ciegciegit	9250
CGTCGCTGGT	TACAGCGGG	GAGACATATY	A TCACAGCCTO	G TCTCGTGCCC	9300
GACCCCCCCTC	GITICCGITC	G TGCCTACTCC	TACTITICIG	r aggggraggc	9350
ATTTACCTGC	TCCCCAACCC	ATGAACGGG	G AGCTAACCA	C TCCAGGCCTT	9400
AAGCCATTIC	CIGITITI	TTTTTTTTT	r tritritit	r TCTTTTTTT	9450
TTTCTTTCC	TICCTICIT	r TTTTCCTTTC	C TITTICCCI	T CTTTAATGGT	9500

	10	20	- 30	40	50	
12	34567890	1234567890	1234567890	1234567890	1234567890	
GC:	CTCCATCT	TAGCCCTAGT	CACGGCTAGC	TGTGAAAGGT	CCCTICACCCC	9550
CA	IGACIGCA	GAGAGICCIG	ATACTGGCCT	CICIGCAGAT	CATGT	9595

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20	30	40	50	
				50
				100
				150
				200
				250
				300
				350
				400
				450
				500
			~	550
				600
				650
				700
				750
				800
			~	850
				900
				950
				1000
				1050
				1100
				1150
			· 	1200
			_	1250
TLGFGAYMSK	AHGIDPNIRT	GVRTTTTGGS	ITYSTYGKFL	1300
DIIICDECHS	TOSTITLGIG	TVLDQAETAG	ARLVVLATAT	1350
				1400
GLNAVAYYRG	LDVSVIPPIG	DVVVVATDAL	MIGFIGDFDS	1450
				1500
				1550
				1600
				1650
				1700
				1750
				1800
				1850
GALVAFKVMS	GEVPSTEDLV	NLLPATLSPG	ALVVGVVCAA	1900
	1234567890 TKRNINRRPQ RRQPIPKARR DPRRSENIG GWYATGNIP SIVYEAADVI HVDLLWGIAA GHVSGHRMAW YYSMWGWAK IQLWNINGSW PIDWFAQGWG PSPWWGITD TKTCOGPPCN YPYRLWHYPC ELSPLILSTT SFALKWEYIL GAHGILSFLW DREMAASCGG VPPLNVRGGR YFVRAQGLIR HAGLRDLAVA FLGPADSLEG WYSTATQSFL WQAPPGARSM LKGSSGGPLL TDNSTPPAVP TLGFGAYMSK DILICDECHS NIEEIGLSNN GLNAVAYYRG TVDFSLDPTF MFDSSVLCEC VFTGLTHIDA LIRLKPTLHG VLVGGVLAAL CASQLPYTEQ WAKHMWNFIS ILGGWVAAQL	1234567890 1234567890 TKRNINRRPQ DVKFPGGQII RRQPIPKARR PEGRAWAQPG DPRRSKNLG KVIDILITOGF GVNYATGNLP GCSFSIFLLA SIVYFAADVI MHIPGCVPCV HVDLLWGIAA FCSAMYVGDL GHVSCHRWW DMMWSPIT YYSMVGNWAK VLIVALLFAG IQLWNINGSW HINRIALNCN PIDWFAQGWG PITYIKPNSS PSPVVVGITD RSGVPIYSWG TKICGGPPCN IGGVGNRILI YPYRLWHYPC TINFSIFKVR ELSPLLLSIT EWQILPCAFT SFAIKWEYIL LLFILLADAR GAHGILSFLV FFCAAWYIKG DREMAASCGG AVLVGLVFLIT VPPLNVRGGR DALILLICAV YFVRAQGLIR ACMLVRKVAG HAGLRDLAVA VEPVVFSAME FIGPADSLEG QGWRLLAPIT VVSIATQSFL ATCINGVCWT WQAPPGARSM TPCSCGSSDL LKGSSGGPLL CPSCHVVGVF TDNSTPPAVP QIFQVAHLHA TLGFGAYMSK AHGIDPNIRT DILICDECHS TDSTTILGIG NIEELGLSNN GEIPFYGKAI GLNAVAYYRG LDVSVIPPIG TVDFSLDPIF TIETTIVPQD MFDSSVLCEC YDAGCAWYEL VFTGLIHIDA HFLSQIKQAG LIRLKPTLHG PTPLLYRLGA VLVGGVLAAL AAYCLITIGSV WAKHMMFIS GIQYLAGLST ILCGWWAAQL APPSAASAFV	TKRVINRRPQ DVKFPGGGQI VGGVYLLPRR RRQPIPKARR PEGRAWAQPG YPWPLYGNEG DPRRRSRNLG KVIDITITOGF ADIMGYIPLV GWYATGALV GGWYATGALV GGSFSIFILA LLSCITIPAS SIVYEAADVI MHIPGCVPCV QEGNESROW HVDLLWGIAA FCSAMYVEDL CGSIFLVGQL GHVSCHWAW VLIVALLFAG VDGEIHTIGR YYSMVCMAK VLIVALLFAG VDGEIHTIGR TQLWINGSW HINRIALNCN DSLQIGFFAA PILWFAQSWG PITYIKPNSS DQRPYOWHYA PSPVWGTID RSGVPIYSWG ENEIDWILLN TKICGGPPCN IGGVENRILI CPIDCFRKHP YPYRLWHYPC TLNFSIFKVR MYVGGVEHRL SFALKWEYIL LLFLLLADAR VCACLWMILL GAHGILSFIV FFCAAWYIKG RLARGAAYAF DREMAASCGG AVLWGLVFLT LSPYYKVFLT VPPLNVRGG DAIILLICAV HPELIFDITK YFVRAQGLIR ACMLVRKVAG GHYVQMVFMK HAGLRDLAVA VEPVVFSAME TKVTIWGADT FLGPADSLEG QGWRLLAPIT AYSQQIRGVL WYKAGSKITL WQAPPGARSM TPCSCGSSDL YLWIRHADVI LKGSSGGPLL CPSCHVVGWF RAAVCTRGVA TINSTPPAVP QIFQVAHLHA PIGSGKSIKV TLGFGAYMSK AHGIDPNIRT GVRTITIGGS NIEBIGLSIN GETPFYGKAI PIFALKOGRH GLNAVAYYRG LDVSVIPPIG DVWVATDAL TVDFSLDPIF TIETITVPQD AVSRQQRGR MFDSSVLCEC YDAGCAWYEL TPAETSVRLR VFIGLIHIDA HFLSQIKQAG INFPYLVAYQ LIRLKPITHG FIPLLYRLGA VQNEVILIHP VLVGGVLAAL AAYCLITGSV VIVGRILLSG CASQLPYTEQ GMQLABQFKQ KALGLLQTAT WAKHMWFIS GIQYLAGLST LPCNPALASL LLGGWAAQL APPSAASAFV GAGGAGAAVG	1234567890 123456780 1234567890 1234567890 123456780 123456780 1234567890 123456780

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
ILRRHVGPGE	GAVQWMNRLI	AFASRONHVS	PIHYVPESDA	AARVIQILSS	1950
LTTTQLLKRL	HOWINEDCST	PCSGSWLRDV	MDWICIVLID	FKIWLQSKLL	2000
PRLPGVPFLS	CORGYKGWR	GDGIMQITCP	CCAQIACHVK	NGSMRIVGPR	2050
		TPSPAPNYSR			2100
YVIGMIIDW	KCPCQVPAPE	FFTEVDGVRL	HRYAPACKPL	LREDVIFQVG	2150
INOYLVGSQL	PCEPEPDVIV	LISMLIDPSH	TTAETAKRRL	ARGSPPSLAS	2200
		SPDADLIEAN			2250
		AAEIIRKSRK			2300
		APPIPPPRRK			2350
				LEGEPGDPDL -	2400
		MSYIWIGALI			2450
RHHNMVYATT	SRSASLRQKK	VIFDRLQVLD	DHYRDVLKEM	KAKASIVKAK	2500
LLSTEEACKL	TPPHSAKSKF	GYGAKDVRNL	SSRAVNHIRS	WEDLLEDIE	2550
		GGRKPARLIV			2600
		EFLVNIWKSK			2650
				QNCGYRRCRA	2700
				VICESAGIQE	2750
DAAALRAFTE	AMIRYSAPPG	DPPQPEYDLE	LITSCSSNVS	VAHDASCKRV	2800
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- Trp Gln Tyr Val Cys Asn Phe Phe Val Ile Cys Phe Asn Val Leu Lys 1875 1880 1885
- Ala Gly Val Gln Ser Met Val Asn Ile Pro Gly Cys Pro Phe Tyr Ser 1890 1895 1900
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Thr Arg Lys Thr Ser Glu Arg Ser Gln Pro Arg Gly Arg Arg Gln Pro 50 55 60

Ile Pro Lys Asp Arg Arg Ser Thr Gly Lys Ser Trp Gly Lys Pro Gly 65 70 75 80

Tyr Pro Trp Pro Leu Tyr Gly Asn Glu Gly Leu Gly Trp Ala Gly Trp
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Leu Leu Ser Pro Arg Gly Ser Arg Pro Ser Trp Gly Pro Asn Asp Pro
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Arg His Arg Ser Arg Asn Val Gly Lys Val Ile Asp Thr Leu Thr Cys 115 120 125

Gly Phe Ala Asp Leu Met Gly Tyr Ile Pro Val Val Gly Ala Pro Leu 130 135 140

Gly Gly Val Ala Arg Ala Leu Ala His Gly Val Arg Val Leu Glu Asp 145 150 155 160

Gly Val Asn Phe Ala Thr Gly Asn Leu Pro Gly Cys Ser Phe Ser Ile 165 170 175

Phe Leu Leu Ala Leu Leu Ser Cys Ile Thr Thr Pro Val Ser Ala Ala

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Glu Val Lys Asn Ile Ser Thr Gly Tyr Met Val Thr Asn Asp Cys Thr 195 200 205

Asn Asp Ser Ile Thr Trp Gln Leu Gln Ala Ala Val Leu His Val Pro 210 215 220

Gly Cys Val Pro Cys Glu Lys Val Gly Asn Ala Ser Gln Cys Trp Ile 225 230 235 240

Pro Val Ser Pro Asn Val Ala Val Gln Arg Pro Gly Ala Leu Thr Gln 245 250 255

Gly Leu Arg Thr His Ile Asp Met Val Val Met Ser Ala Thr Leu Cys 260 265 270

Ser Ala Leu Tyr Val Gly Asp Leu Cys Gly Gly Val Met Leu Ala Ala 275 280 285

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Asp Met Met Met Asn Trp Ser Pro Thr Ala Thr Met Ile Leu Ala Tyr 325 330 335

Ala Met Arg Val Pro Glu Val Ile Ile Asp Ile Ile Ser Gly Ala His 340 345 350

Trp Gly Val Met Phe Gly Leu Ala Tyr Phe Ser Met Gln Gly Ala Trp 355 360 365

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Thr His Thr Val Gly Gly Ser Ala Ala Gln Thr Thr Gly Arg Leu Thr 385 390 395 400

Ser Leu Phe Asp Met Gly Pro Arg Gln Lys Ile Gln Leu Val Asn Thr 405 410 415

Asn Gly Ser Trp His Ile Asn Arg Thr Ala Leu Asn Cys Asn Asp Ser
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Pro Ser Pro Val Val Val Gly Thr Thr Asp Arg Leu Gly Ala Pro Thr

Tyr Thr Trp Gly Glu Asn Glu Thr Asp Val Phe Leu Leu Asn Ser Thr

Arg Pro Pro Leu Gly Ser Trp Phe Gly Cys Thr Trp Met Asn Ser Ser

Gly Tyr Thr Lys Thr Cys Gly Ala Pro Pro Cys Arg Thr Arg Ala Asp

Phe Asn Ala Ser Thr Asp Leu Leu Cys Pro Thr Asp Cys Phe Arg Lys

His Pro Asp Thr Thr Tyr Leu Lys Cys Gly Ser Gly Pro Trp Leu Thr

Pro Arg Cys Leu Ile Asp Tyr Pro Tyr Arg Leu Trp His Tyr Pro Cys

Thr Val Asn Tyr Thr Ile Phe Lys Ile Arg Met Tyr Val Gly Gly Val

Glu His Arg Leu Thr Ala Ala Cys Asn Phe Thr Arg Gly Asp Arg Cys

Asn Leu Glu Asp Arg Asp Arg Ser Gln Leu Ser Pro Leu Leu His Ser

Thr Thr Glu Trp Ala Ile Leu Pro Cys Ser Tyr Ser Asp Leu Pro Ala

Leu Ser Thr Gly Leu Leu His Leu His Gln Asn Ile Val Asp Val Gln

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Phe Met Tyr Gly Leu Ser Pro Ala Leu Thr Lys Tyr Ile Val Arg Trp 705 710 715 720

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Ala Cys Leu Trp Met Leu Ile Leu Leu Gly Gln Ala Glu Ala Ala Leu
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Glu Lys Leu Val Ile Leu His Ala Ala Ser Ala Ala Ser Cys Asn Gly
755 760 765

Phe Leu Tyr Phe Val Ile Phe Phe Val Ala Ala Trp Tyr Ile Lys Gly
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Arg Val Val Pro Leu Ala Thr Tyr Ser Leu Thr Gly Leu Trp Ser Phe 785 790 795 800

Ser Leu Leu Leu Ala Leu Pro Gln Gln Ala Tyr Ala Tyr Asp Ala 805 810 815

Ser Val His Gly Gln Ile Gly Ala Ala Leu Leu Val Met Ile Thr Leu 820 825 830

Phe Thr Leu Thr Pro Gly Tyr Lys Thr Leu Leu Ser Arg Phe Leu Trp 835 840 845

Trp Leu Cys Tyr Leu Leu Thr Leu Gly Glu Ala Met Val Gln Glu Trp 850 855 860

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Val Pro Tyr Phe Val Arg Ala His Ala Leu Leu Arg Met Cys Thr Met 915 920 925

Ala Arg His Leu Ala Gly Gly Arg Tyr Val Gln Met Ala Leu Leu Ala 930 935 940

Leu Gly Arg Trp Thr Gly Thr Tyr Ile Tyr Asp His Leu Thr Pro Met

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- Ser Glu Thr Thr Val Arg Leu Arg Ala Tyr Phe Asn Thr Pro Gly Leu 1540 1545 1550
- Pro Val Cys Gln Asp His Leu Glu Phe Trp Glu Ala Val Phe Thr Gly 1555 1560 1565
- Leu Thr His Ile Asp Ala His Phe Leu Ser Gln Thr Lys Gln Ser Gly 1570 1575 1580
- Glu Asn Phe Ala Tyr Leu Thr Ala Tyr Gln Ala Thr Val Cys Ala Arg 1585 1590 1595 1600
- Ala Lys Ala Pro Pro Pro Ser Trp Asp Val Met Trp Lys Cys Leu Thr 1605 1610 1615
- Arg Leu Lys Pro Thr Leu Val Gly Pro Thr Pro Leu Leu Tyr Arg Leu 1620 1625 1630
- Gly Ser Val Thr Asn Glu Val Thr Leu Thr His Pro Val Thr Lys Tyr 1635 1640 1645
- Ile Ala Thr Cys Met Gln Ala Asp Leu Glu Val Met Thr Ser Thr Trp 1650 1655 1660
- Val Leu Ala Gly Gly Val Leu Ala Ala Val Ala Ala Tyr Cys Leu Ala 1665 1670 1675 1680
- Thr Gly Cys Val Cys Ile Ile Gly Arg Leu His Ile Asn Gln Arg Ala 1685 1690 1695
- Val Val Ala Pro Asp Lys Glu Val Leu Tyr Glu Ala Phe Asp Glu Met 1700 1705 1710
- Glu Glu Cys Ala Ser Arg Ala Ala Leu Ile Glu Glu Gly Gln Arg Ile

1715 1720 1725

Ala Glu Met Leu Lys Ser Lys Ile Gln Gly Leu Leu Gln Gln Ala Ser 1730 1735 1740

- Lys Gln Ala Gln Asp Ile Gln Pro Thr Val Gln Ala Ser Trp Pro Lys 1745 1750 1755 1760
- Val Glu Gln Phe Trp Ala Lys His Met Trp Asn Phe Ile Ser Gly Ile 1765 1770 1775
- Gln Tyr Leu Ala Gly Leu Ser Thr Leu Pro Gly Asn Pro Ala Val Ala 1780 1785 1790
- Ser Met Met Ala Phe Ser Ala Ala Leu Thr Ser Pro Leu Ser Thr Ser 1795 1800 1805
- Thr Thr Ile Leu Leu Asn Ile Leu Gly Gly Trp Leu Ala Ser Gln Ile 1810 1815 1820
- Ala Pro Pro Ala Gly Ala Thr Gly Phe Val Val Ser Gly Leu Val Gly 1825 1830 1835 1840
- Ala Ala Val Gly Ser Ile Gly Leu Gly Lys Val Leu Val Asp Ile Leu 1845 1850 1855
- Ala Gly Tyr Gly Ala Gly Ile Ser Gly Ala Leu Val Ala Phe Lys Ile 1860 1865 1870
- Met Ser Gly Glu Lys Pro Ser Met Glu Asp Val Val Asn Leu Leu Pro 1875 1880 1885
- Gly Ile Leu Ser Pro Gly Ala Leu Val Val Gly Val Ile Cys Ala Ala 1890 1895 1900
- Ile Leu Arg Arg His Val Gly Pro Gly Glu Gly Ala Val Gln Trp Met 1905 1910 1915 1920
- Asn Arg Leu Ile Ala Phe Ala Ser Arg Gly Asn His Val Ala Pro Thr 1925 1930 1935
- His Tyr Val Thr Glu Ser Asp Ala Ser Gln Arg Val Thr Gln Leu Leu 1940 1945 1950
- Gly Ser Leu Thr Ile Thr Ser Leu Leu Arg Arg Leu His Asn Trp Ile 1955 1960 1965
- Thr Glu Asp Cys Pro Ile Pro Cys Gly Gly Ser Trp Leu Arg Asp Val

1970 1975 1980

Trp Asp Trp Val Cys Thr Ile Leu Thr Asp Phe Lys Asn Trp Leu Thr 1985 1990 1995 2000

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- Ser Lys Leu Phe Pro Lys Met Pro Gly Leu Pro Phe Val Ser Cys Gln 2005 2010 2015
- Lys Gly Tyr Lys Gly Val Trp Ala Gly Thr Gly Ile Met Thr Thr Arg 2020 2025 2030
- Cys Pro Cys Gly Ala Asn Ile Ser Gly Asn Val Arg Leu Gly Ser Met 2035 2040 2045
- Arg Ile Thr Gly Pro Lys Thr Cys Met Asn Ile Trp Gln Gly Thr Phe 2050 2055 2060
- Pro Ile Asn Cys Tyr Thr Glu Gly Gln Cys Val Pro Lys Pro Ala Pro 2065 2070 2075 2080
- Asn Phe Lys Val Ala Ile Trp Arg Val Ala Ala Ser Glu Tyr Ala Glu 2085 2090 2095
- Val Thr Gln His Gly Ser Tyr His Tyr Ile Thr Gly Leu Thr Thr Asp 2100 2105 2110
- Asn Leu Lys Val Pro Cys Gln Leu Pro Ser Pro Glu Phe Phe Ser Trp 2115 2120 2125
- Val Asp Gly Val Gln Ile His Arg Phe Ala Pro Thr Pro Lys Pro Phe 2130 2135 2140
- Phe Arg Asp Glu Val Ser Phe Cys Val Gly Leu Asn Ser Phe Val Val 2145 2150 2155 2160
- Gly Ser Gln Leu Pro Cys Asp Pro Glu Pro Asp Thr Asp Val Leu Met \$2165\$ \$2170\$ \$2175\$
- Ser Met Leu Thr Asp Pro Ser His Ile Thr Ala Glu Thr Ala Ala Arg 2180 2185 2190
- Arg Leu Ala Arg Gly Ser Pro Pro Ser Glu Ala Ser Ser Ser Ala Ser 2195 2200 2205
- Gln Leu Ser Ala Pro Ser Leu Arg Ala Thr Cys Thr Thr His Gly Lys 2210 2215 2220
- Ala Tyr Asp Val Asp Met Val Asp Ala Asn Leu Phe Met Gly Gly Asp

2225 2230 2235 2240

Val Thr Arg Ile Glu Ser Gly Ser Lys Val Val Leu Asp Ser Leu 2245 2250 2255

- Asp Pro Met Val Glu Glu Arg Ser Asp Leu Glu Pro Ser Ile Pro Ser 2260 2265 2270
- Glu Tyr Met Leu Pro Lys Lys Arg Phe Pro Pro Ala Leu Pro Ala Trp 2275 2280 2285
- Ala Arg Pro Asp Tyr Asn Pro Pro Leu Val Glu Ser Trp Lys Arg Pro 2290 2295 2300
- Asp Tyr Gln Pro Ala Thr Val Ala Gly Cys Ala Leu Pro Pro Pro Arg 2305 2310 2315 2320
- Lys Thr Pro Thr Pro Pro Pro Arg Arg Arg Thr Val Gly Leu Ser 2325 2330 2335
- Glu Asp Ser Ile Gly Asp Ala Leu Gln Gln Leu Ala Ile Lys Ser Phe 2340 2345 2350
- Gly Gln Pro Pro Pro Ser Gly Asp Ser Gly Leu Ser Thr Gly Ala Gly
 2355 2360 2365
- Ala Ala Asp Ser Gly Ser Gln Thr Pro Pro Asp Glu Leu Ala Leu Ser 2370 2375 2380
- Glu Thr Gly Ser Ile Ser Ser Met Pro Pro Leu Glu Gly Glu Leu Gly 2385 2390 2395 2400
- Asp Pro Asp Leu Glu Pro Glu Gln Val Glu Pro Gln Pro Pro Pro Gln 2405 2410 2415
- Gly Gly Val Ala Ala Pro Gly Ser Asp Ser Gly Ser Trp Ser Thr Cys 2420 2425 2430
- Ser Glu Glu Asp Asp Ser Val Val Cys Cys Ser Met Ser Tyr Ser Trp 2435 2440 2445
- Thr Gly Ala Leu Ile Thr Pro Cys Ser Pro Glu Glu Glu Lys Leu Pro 2450 2455 2460
- Ile Asn Pro Leu Ser Asn Ser Leu Leu Arg Tyr His Asn Lys Val Tyr 2465 2470 2475 2480
- Cys Thr Thr Lys Ser Ala Ser Leu Arg Ala Lys Lys Val Thr Phe

2485 2490 2495

- Asp Arg Met Gln Val Leu Asp Ser Tyr Tyr Asp Ser Val Leu Lys Asp 2500 2505 2510
- Ile Lys Leu Ala Ala Ser Lys Val Thr Ala Arg Leu Leu Thr Met Glu 2515 2520 2525
- Glu Ala Cys Gln Leu Thr Pro Pro His Ser Ala Arg Ser Lys Tyr Gly 2530 2535 2540
- Phe Gly Ala Lys Glu Val Arg Ser Leu Ser Gly Arg Ala Val Asn His 2545 2550 2555 2560
- Ile Lys Ser Val Trp Lys Asp Leu Leu Glu Asp Ser Glu Thr Pro Ile
 2565 2570 2575
- Pro Thr Thr Ile Met Ala Lys Asn Glu Val Phe Cys Val Asp Pro Thr 2580 2585 2590
- Lys Gly Gly Lys Lys Ala Ala Arg Leu Ile Val Tyr Pro Asp Leu Gly 2595 2600 2605
- Val Arg Val Cys Glu Lys Met Ala Leu Tyr Asp Ile Thr Gln Lys Leu 2610 2615 2620
- Pro Gln Ala Val Met Gly Ala Ser Tyr Gly Phe Gln Tyr Ser Pro Ala 2625 2630 2635 2640
- Gln Arg Val Glu Phe Leu Leu Lys Ala Trp Ala Glu Lys Lys Asp Pro 2645 2650 2655
- Met Gly Phe Ser Tyr Asp Thr Arg Cys Phe Asp Ser Thr Val Thr Glu 2660 2665 2670
- Arg Asp Ile Arg Thr Glu Glu Ser Ile Tyr Arg Ala Cys Ser Leu Pro 2675 2680 2685
- Glu Glu Ala His Thr Ala Ile His Ser Leu Thr Glu Arg Leu Tyr Val 2690 2695 2700
- Gly Gly Pro Met Phe Asn Ser Lys Gly Gln Thr Cys Gly Tyr Arg Arg 2705 2710 2715 2720
- Cys Arg Ala Ser Gly Val Leu Thr Thr Ser Met Gly Asn Thr Ile Thr 2725 2730 2735
- Cys Tyr Val Lys Ala Leu Ala Ala Cys Lys Ala Ala Gly Ile Ile Ala

2740 2745 2750

Pro Thr Met Leu Val Cys Gly Asp Asp Leu Val Val Ile Ser Glu Ser 2755 2760 2765

Gln Gly Thr Glu Glu Asp Glu Arg Asn Leu Arg Ala Phe Thr Glu Ala 2770 2775 2780

Met Thr Arg Tyr Ser Ala Pro Pro Gly Asp Pro Pro Arg Pro Glu Tyr 2785 2790 2795 2800

Asp Leu Glu Leu Ile Thr Ser Cys Ser Ser Asn Val Ser Val Ala Leu 2805 2810 2815

Gly Pro Gln Gly Arg Arg Tyr Tyr Leu Thr Arg Asp Pro Thr Thr 2820 2825 2830

Pro Ile Ala Arg Ala Ala Trp Glu Thr Val Arg His Ser Pro Val Asn 2835 2840 2845

Ser Trp Leu Gly Asn Ile Ile Gln Tyr Ala Pro Thr Ile Trp Ala Arg 2850 2855 2860

Met Val Leu Met Thr His Phe Phe Ser Ile Leu Met Ala Gln Asp Thr 2865 2870 2875 2880

Leu Asp Gln Asn Leu Asn Phe Glu Met Tyr Gly Ala Val Tyr Ser Val 2885 2890 2895

Ser Pro Leu Asp Leu Pro Ala Ile Ile Glu Arg Leu His Gly Leu Asp 2900 2905 2910

Ala Phe Ser Leu His Thr Tyr Thr Pro His Glu Leu Thr Arg Val Ala 2915 2920 2925

Ser Ala Leu Arg Lys Leu Gly Ala Pro Pro Leu Arg Ala Trp Lys Ser 2930 2935 2940

Arg Ala Arg Ala Val Arg Ala Ser Leu Ile Ser Arg Gly Gly Arg Ala 2945 2950 2955 2960

Ala Val Cys Gly Arg Tyr Leu Phe Asn Trp Ala Val Lys Thr Lys Leu 2965 2970 2975

Lys Leu Thr Pro Leu Pro Glu Ala Arg Leu Leu Asp Leu Ser Ser Trp 2980 2985 2990

Phe Thr Val Gly Ala Gly Gly Gly Asp Ile Tyr His Ser Val Ser Arg

2995 3000 3005

Ala Arg Pro Arg Leu Leu Leu Phe Gly Leu Leu Leu Leu Phe Val Gly 3010 3015 3020

Val Gly Leu Phe Leu Leu Pro Ala Arg 3025 3030

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A. CLASSI IPC 7	FICATION OF SUBJECT MATTER C12N15/51 C07K14/18 C12Q1	/68 C12N7/00	
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Documental	tion searched other than minimum documentation to the extent t	that such documents are included in the fields s	earched
Electronic d	ata base consulted during the international search (name of da	ta base and, where practical, search terms used	i)
EPO-In	ternal, BIOSIS, MEDLINE		
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X	WO 95 21922 A (PILOT MATIAS TA SHERI L (US); SIMONS JOHN N (U 17 August 1995 (1995-08-17) page 4, line 18 -page 6, line page 55, line 24 -page 56, line page 76; example 5 page 89, line 18 -page 96 page 109; example 15 page 148; example 21 page 427, line 17 -page 432 claims	S); ABBOT) 17	1,2,4-18
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Date of the	actual completion of the international search	Date of mailing of the international se	earch report
	7 October 2000	31/10/2000	
Name and r	mailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fav. (+31-70) 340-3018	Authorized officer Andres, S	

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